Multiple species of morels occur in landscape settings

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Abstract: Morels (*Morchella* spp.) are popular ascomycete fungi due to their culinary value. Recent advancements in molecular identification have spurred interest in the ecology and genetics of these edible fungi. Most morels are known to inhabit forest settings and may have mycorrhizal associations with trees. Two species of morels, M. importuna and M. rufobrunnea, are associated with urban landscape settings and M. rufobrunnea has been cultivated, suggesting that it has a saprotrophic nutritional mode. As part of a graduate class project at Washington State University, morels collected from landscape settings in the Pacific Northwest of the United States and British Columbia, Canada were identified using sequence data. Five phylogenetic species were identified and two species have not been previously reported from urban settings. Of the three species previously reported from landscape settings, one represents the first report of this species from a North American landscape setting, and another is only the third specimen collected of an as of yet unnamed

species. Our findings suggest that a diverse range of morels occur in landscape and urban settings.

Key words: Canada, distribution, ecology, *Morchellaceae*, mulch morel, United States.

Introduction

Morels (*Morchella* spp.) are some of the most highly prized edible macrofungi. Morels are thought to have originated in the Holarctic over 120 million years ago and are now distributed worldwide (O'Donnell et al., 2011). Nineteen phylogenetic lineages of North American morels identified by O'Donnell et al. (2011) were formally described as species by Kuo et al. (2012). More recently, Richard et al. (2015) reconciled the North American species described by Kuo et al. (2012) with those described by Clowez (2010). The majority of North American species has been reported from forested and other "non-urban" environments and may have mycorrhizal associations with trees (Dahlstrom et al., 2000).



▼ o-called "landscape" or "mulch morels" are of particular interest and have been identified and characterized based on their ecological niche (Kuo et al., 2012; Mann and Mann, 2014; Ondřej et al., 2011). Two species of morels, M. rufobrunnea and *M. importuna*, are the most common species inhabiting landscape settings (Guzmán and Tapia, 1998; Kuo, 2008; Kuo et al., 2012; Mann and Mann, 2014). Such landscapes include bark or mulched beds, gardens, planters, lawns, and roadsides (Guzmán and Tapia, 1998; Kuo, 2008; O'Donnell et al., 2011). Many morels are found in association with trees (Kuo et al., 2012) and are suspected to form mycorrhizal relationships (Dahlstrom et al., 2000), however, both *M. rufobrunnea* and *M. importuna* are suspected saprotrophs due to their profundity in landscapes and apparent lack of association with trees (Mann and Mann, 2014). Despite the reported prevalence of these two species

from urban environments (Guzmán and Tapia, 1998; Kuo, 2008; Kuo et al., 2012; Mann and Mann, 2014), a few morel species have been found to inhabit both landscape and forest settings (Kuo et al., 2012; Ondřej et al., 2011). As part of a spring 2014 class project in "Advanced Fungal Biology," a graduate-level plant pathology course at Washington State University (WSU), we characterized specimens of morels collected from landscape and urban settings using DNA-sequence data. Our aim was to determine the diversity and ecology of landscape morels present at several sites in Washington State, Idaho, and British Columbia.

Materials and Methods

Morel fruiting bodies were collected from April to May 2011-2013 from landscape settings including lawns, landscaping bark and rocks, and mulched plant beds in Washington, Idaho, and British Columbia. Specimens were placed on paper overnight at room

temperature (20-25°C) to generate spore prints, and then allowed to dry at room temperature prior to storage. Voucher specimens were deposited in WSU's Charles Gardner Shaw Mycological Herbarium (accessions in Fig. 1). Single-ascospore cultures were grown on potato dextrose agar and (PDA) (BD, Franklin Lakes, New Jersey) amended with penicillin (500 units/L) and streptomycin (500 µg/L) to inhibit bacterial contamination. If the specimen lacked mature ascospores, a culture was initiated from surface sterilized stipe tissue. Subcultures for DNA extraction were grown on sterile dialysis membrane (SpectraPor, VWR Scientific, Radnor, Pennsylvania) overlaying PDA amended with antibiotics (BD, Franklin Lakes, New Jersey). Mycelia were gently scraped from the membranes after 5-7 days, frozen, and lyophilized. DNA extraction, PCR amplification, sequencing, and phylogenetic analysis followed the methods described in

Those pursuing collection of morels for the table from landscape and urban settings should proceed with caution. Morels and other mushrooms are known bioaccumulators of heavy metals (Cocci et al., 2006). A study published in FUNGI in 2010 showed arsenic and lead accumulation in morels collected in abandoned apple orchards previously treated with commercial pesticides (Shavit and Shavit, 2010). Furthermore, there is at least one report of a morel fruiting following an application of the herbicide atrazine (Goldway et al., 2000). Thus it is advisable to consume landscape morels sparingly or not at all unless one has intimate knowledge of the history of the fruiting site including any past chemical applications or toxic contamination due to the potential for contamination of the fruiting body.

Carris et al. (2015). Four loci were used: translation elongation factor $1-\alpha$ (EF1- α), the 28s ribosomal subunit (28s), RNA polymerase II largest subunit (RPB1), and RNA polymerase II second largest subunit (RPB2). Representative sequences were deposited in GenBank (accessions in caption of Fig. 1 – page 30).

Results and Conclusions

Our analysis reveals that the diversity of morel species occurring in landscape settings is likely much greater than is currently recognized. Eleven specimens of morels collected from landscape



settings belonged to five different phylogenetic species of morels (Fig. 1) based on the results of our phylogenetic analysis. Seven of the specimens, collected from Salt Spring Island, British Columbia, Moscow, ID, and Pullman, WA, were identified as *M. importuna*. Morchella importuna was collected from different types of landscapes including landscape gravel, mulch, and a grassy lawn. Other specimens were identified as M. brunnea, M. populiphila, M. snyderi, and an unnamed species corresponding to phylogenetic lineage Mel-8 (Kuo et al., 2012; O'Donnell et al., 2011). Specimens of M. brunnea and *M. populiphila* were characterized

as part of another study focusing on the occurrence of the asexual stage of morels and false morels (Carris et al., 2015), and represented the first report of M. brunnea and M. populiphila from landscape settings. Morchella brunnea was described by Kuo et al. (2012) as occurring under hardwoods (Acer and Quercus) and likely also occurs in non-burned conifer forests in the Pacific Northwest. The M. brunnea collections in this study were fruiting in landscape bark and an adjacent lawn in a residential yard in Pullman, WA (Carris et al., 2015). Nearby woody plants included species of

Continued on page 32.



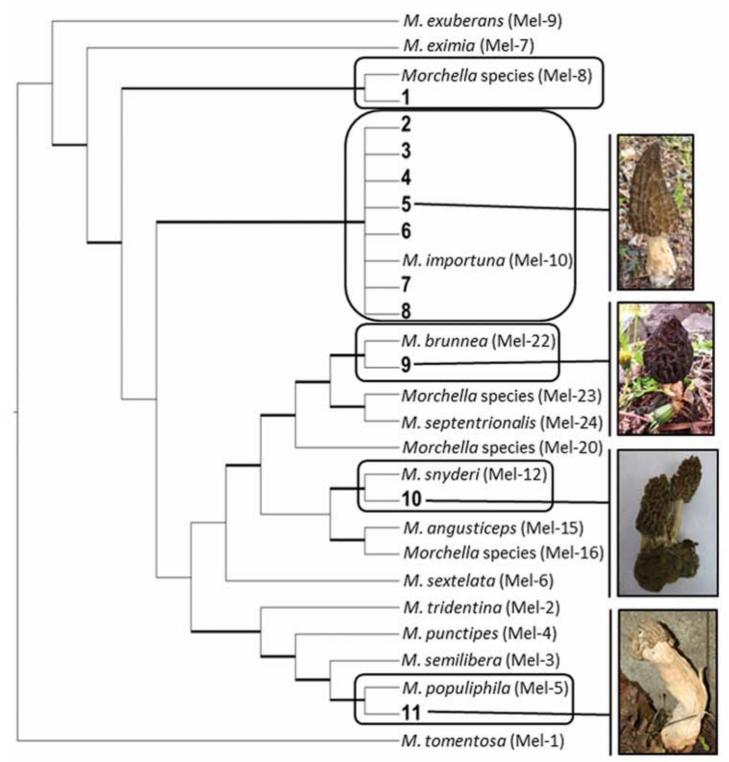


Figure 1. Cladogram describing phylogenetic relationship among various morel species from the *M. elata* clade (Kuo et al., 2012; O'Donnell et al., 2011) and landscape morels reported in this study with *M. tomentosa* as the outgroup. Five clades (circled) are represented from specimens genotyped in this study. Specimens, designated by numbers from 1 through 11, were collected from landscape settings in: 1. WSP 72734, Pullman, WA, USA (landscape bark mulch); 2. WSP 72735, Salt Spring Island, BC, Canada (landscape gravel); 3. WSP 72733, Moscow, ID, USA (landscape bark mulch); 4-5. WSP 72736 and 72737, Salt Spring Island, BC, Canada (landscape gravel); 6. WSP 72356, Pullman, WA, USA (WSU Food Science and Human Nutrition Building, landscape bark mulch). 7. WSP 72732, Pullman, WA, USA (WSU Campus Police Station, landscape bark mulch). 8.

Pullman, WA, USA (WSU Compton Union Building, landscape bark mulc

Union Building, landscape bark mulch; specimen not available). 9. WSP 72719, Pullman, WA, USA (residential yard, landscape bark mulch). 10. WSP 72738, Pullman, WA, USA (WSU Ensminger Pavilion, landscape bark mulch). 11. WSP 72720, Pullman, WA, USA (landscape bark mulch and lawn). Photos of selected specimens are included. Bolded lines represent bootstrap values and posterior probabilities greater than 70 and 95, respectively. Unique sequences from this study were deposited in GenBank under accession numbers KP842238-KP842261. Sequences for specimen numbers 6, 9 and 11 were deposited in GenBank separately by Carris et al. (2015). Supplementary sequences were obtained from Kuo et al. (2012), O'Donnell et al. (2011) and, Taşkin et al. (2012).



Mel-8 is considered a "black morel," a group distinguished by the presence of a vallécula, where the bottom of the cap hangs slightly over the stipe (Bunyard, 2013). Ridges on this species are vertically-aligned on a conical-shaped, slightly rounded cap. It is difficult to distinguish this species from M. brunnea based on morphology alone.



M. importuna is considered the "typical" landscape morel as it seems to be the most common in landscape settings. Also a "black morel," this morel can be recognized by its vertically-arranged, deep pits with ladder-like, horizontally-aligned ridges.



M. brunnea is also a "black morel" and closely resembles Mel-8 and its eastern U.S. look-alike. M. angusticeps. The cap is nearly conical with vertically-aligned pits, however, without the ladderlike ridges of M. importuna



M. snyderi can be easily confused with the "yellow morel" due to its pale color when immature, however, yellow morels do not have a vallécula which is present in M. snyderi. This morel turns dark brown as it matures and has a characteristic stipe which becomes increasingly ridged and pocketed with age.



M. populiphila has only been recently distinguished from its look-alike, M. semilibera or the "half-free morel," due to its cap that overhangs significantly over the stipe without attachment. When young, the cap is pale colored, but matures dark brown to black.





al., 2011). Interestingly, the unnamed species (Mel-8) was also collected from the WSU Ensminger Pavilion site later in the season, indicating that at least two different species were fruiting in the same landscape setting. The WSU Ensminger Pavilion specimen is the third time this unnamed species has been collected. The first collection was made in 1990 by Nancy Smith Weber at a post-fire forest site in Oregon (specimen NSW 6190; O'Donnell et al., 2011; Kuo et al., 2012). The second collection of Mel-8 was under an incense cedar in an old apple orchard in a northern California backyard (see "Morchella species Mel-8" MushroomExpert. com). Little can be concluded about the ecology of this species with so few reported findings, but the diversity of habitats from which the three known specimens were collected is intriguing. As more genetic data become available for landscape morel specimens, it will be interesting to see if the distinction between urban and forest morels continues to break down, as suggested by the diversity of species observed in this study.

Acknowledgements

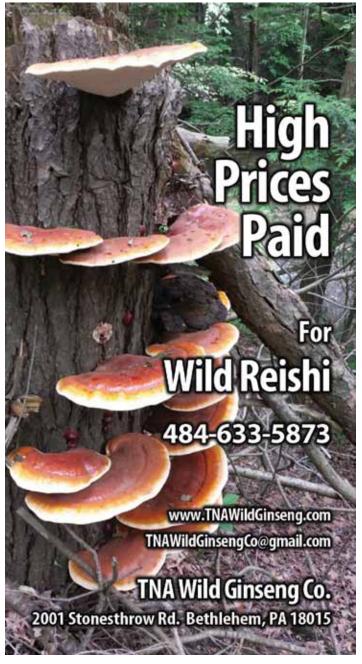
We would like to thank Igbal Aujla, Chongjing Xia, and Sean Mullahy for their help in the culture of morels for this project. We would also like to thank Axel Elling, David Wheeler, and Jeremiah Dung for collection of mulch morels. Thank you also to Dr. Kerry O'Donnell for his advice and information on our Mel-8 specimen. Lastly, we would like to thank Dr. Frank Dugan for reviewing this manuscript prior to submission.

PPNS #0687, Department of Plant Pathology, College of Agricultural, Human, and Natural Resource Sciences, Agricultural Research Center, Hatch Project No. WNP00687, Washington State University, Pullman, WA 99164-6430, USA.

Literature Cited

Bunyard, B. 2013. Morels: the name game. *Fungi* 6(1): 27-32. Carris, L.M., T.L. Peever, and S.W. McCotter. 2015. Mitosporic stages of Disciotis, Gyromitra and Morchella in the inland Pacific Northwestern U.S.A. Mycologia (in press).

Clowez, P. 2010. Les Morilles. Une nouvelle approche mondiale due genre Morchella. Bulletin de la Société Mycologique de France. 126: 199-376.



Cocchi, L., L. Vescovi, L.E. Petrini, and O. Petrini. 2006. Heavy metals in edible mushrooms in Italy. *Food Chemistry* 98: 278–284, doi:10.1016/j.foodchem.2005.05.068.

Dahlstrom, J.L., J.E. Smith, and N.S. Weber. 2000. Mycorrhiza-like interaction by *Morchella* with species of the Pinaceae in pure culture synthesis. *Mycorrhiza* 9: 279–285, doi:10.1007/PL00009992.

Goldway, M., R. Amir, D. Goldberg, Y. Hadar, and D. Levanon. 2000. *Morchella conica* exhibiting a long fruiting season. *Mycological Research* 104(8): 1000–1004.

Guzmán, G., and F. Tapia. 1998. The known morels in Mexico, a description of a new blushing species, *Morchella rufobrunnea*, and new data on *M. guatemalensis*. *Mycologia* 90: 705–714, doi:10.2307/3761230.

Kuo, M. 2008. *Morchella tomentosa*, a new species from western North America, and notes on *M. rufobrunnea*. *Mycotaxon* 105: 441–446.

Kuo, M., D.R. Dewsbury, C.M. Carter, S.A. Rehner, J.D. Moore, J.M. Moncalvo, S.A. Canfield, S.L. Stephenson, A.S. Methven, and T.J. Volk. 2012. Taxonomic revision of true morels (*Morchella*) in Canada and the United States. *Mycologia* 104(5): 1159–1177.

Mann, H., and P. Mann. 2014. *Morchella importuna*: the Pasadena mulch morel. *Omphalina* 5: 11–12.

O'Donnell, K., A.P. Rooney, G.L. Mills, M. Kuo, N.S. Weber, and S.A. Rehner. 2011. Phylogeny and historical biogeography of true morels (*Morchella*) reveals an early Cretaceous origin

and high continental endemism and provincialism in the Holarctic. *Fungal Genetics and Biology* 48: 252–265.

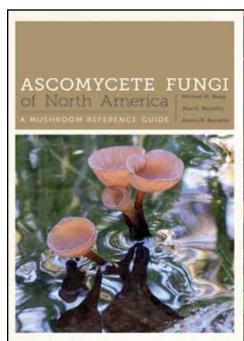
Ondřej, V., P. Havranek, M. Kitner, and P. Nemcova. 2011. Molecular identification and characterization of the edible and medicinal Morchellaceae germplasm collection of "mulch morels." *International Journal of Medicinal Mushrooms* 13: 369–375.

Pagliaccia, D., G.W. Douhan, L. Douhan, T.L. Peever, L.M. Carris, J.L. Kerrigan. 2011. Development of molecular markers and preliminary investigation of the population structure and mating system in one lineage of black morel (*Morchella elata*) in the Pacific Northwestern USA. *Mycologia* 103(5): 969–982, doi:10.3852/10-384.

Richard, F., M. Sauve, J.-M. Bellanger, P. Clowez, K. Hansen, K. O'Donnell, A. Urban, R. Courtecuisse, and P.-A. Moreau. 2015. True morels (*Morchella*, Pezizales) of Europe and North America: evolutionary relationships inferred from multilocus data and a unified taxonomy. *Mycologia* 107(2): 359-382, doi: 10.3852/14-166.

Shavit, E., and E. Shavit. 2010. Lead and arsenic in *Morchella esculenta* fruitbodies collected in lead arsenate contaminated apple orchards in the northeastern United States: A preliminary study. *Fungi* 3(2): 19-26.

Taşkin, H., S. Büyükalaca, K. Hansen, and K. O'Donnell. 2012. Multilocus phylogenetic analysis of true morels (*Morchella*) reveals high levels of endemics in Turkey relative to other regions of Europe. *Mycologia* 104(2): 446−461. ■





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